# Consequences of Host Plant Chemical and Physical Variability to an Associated Herbivore

Gwendolyn L. Waring,\* Department of Biological Sciences, Northern Arizona University, Flagstaff, Arizona 86011, U.S.A.

and

Peter W. Price, Department of Biological Sciences, Northern Arizona University, Flagstaff, Arizona 86011, U.S.A.

#### Abstract

The effects of watering and fertilizer treatments on the vigor and biochemistry of the willow, *Salix lasiolepis*, and subsequent colonization and survivorship of its gallforming herbivore, *Euura lasiolepis*, were investigated in two field experiments. Some plants received low (LW), intermediate (MW) or high (HW) levels of water as treatments, while others received no (OF), low (LF) or high (HF) fertilizer levels. In the watering experiment, plant protein concentrations decreased, while growth rate and number of galls per plant increased with increased water treatments. Plant growth proved to be the best correlate of sawfly attack. Sawfly survivorship increased slightly with greater watering, and phenol concentrations showed no pattern among treatments. In the fertilization experiment, leaf protein increased with fertilization, although shoot length, number of galls and survivorship of *E. lasiolepis* survivorship were greatest in intermediate treatment plants. In both experiments, plant growth, rather than protein or phenol levels, was the best predictor of sawfly attack and survivorship.

In a natural experiment with galls on wild plants, galled tissue had significantly greater protein concentrations and lower phenol concentrations than did ungalled tissue. We suggest that gallformers modify host plant biochemistry within willow galls, which may explain why the chemical parameters of ambient plant quality we tested were less predictive than plant growth.

Key words: *Euura lasiolepis*; Fertilizer experiment; Gallformers; Herbivores; Herbivory; *Salix lasiolepis*; Survivorship; Water experiment.

#### Introduction

Physical and chemical characteristics are the two plant features that herbivores interact with. The chemical basis of plant-herbivore interactions has been increasingly emphasized since the concept of coevolutionary responses to plant chemical defense has become established (Fraenkel, 1959, 1969; Ehrlich and Raven, 1964), and extensively reviewed (e.g. Harborne, 1972, 1977, 1978; Wallace and Mansell, 1976; Chapman and Bernays, 1978; Rosenthal and Janzen, 1979; Visser and Minks, 1982; Hedin, 1983). However, the physical

Accepted August 25, 1988

<sup>\*</sup>Present address: Department of Biology, Museum of Northern Arizona, Route 4, Box 720, Flagstaff, AZ 86001, U.S.A.

attributes of plants, such as growth and structural characteristics, and their relation to herbivory have received much less attention. Because physical properties of plants have chemical bases in terms of growth regulation and chemical constituents of plant parts, physical and chemical properties may be closely correlated. Therefore, we studied relationships between physical and chemical traits in a plant species and their relative influence on host plant selection and survival of an insect herbivore.

An emphasis of plant-herbivore ecology has been that stressed plants are superior resources for herbivores because nutritional quality is improved, plant defenses are reduced, or both (e.g. White, 1974, 1976, 1978, 1984; Rhoades, 1979, 1983; Berryman, 1982; Cates et al., 1983). While the evidence is compelling, these mechanisms are not universal; many herbivores become most abundant on vigorous plants. Van Emden et al. (1969) cite many cases in which fertilizer treatments to plants increase aphid populations. Tetranychid mites frequently respond similarly (Huffaker et al., 1969; Hussey and Huffaker, 1976). Fertilizer treatments to trees increase herbivore numbers in some cases (Bakke, 1969; Shaw and Little, 1972; Onuff et al., 1977).

Plant vigor is important for many plant parasites such as gall formers, as they attack meristematic tissue (Weis et al., 1988). Growth rates of dwarf mistletoes are correlated positively with host plant growth rates (Hawksworth, 1960). Washburn and Cornell (1981) reported on a brief epidemic of the gall wasp, *Xanthoteras politum* (Bassett), that occurred on rapidly growing oak shoots sprouting after a fire. Wasp densities declined within 3 years as plants aged and exhibited reduced growth. Frankie and Morgan (1984) have shown that young oak trees that are susceptible to the gall wasp, *Disholcaspis cinerosa* (Bassett), become resistant as the trees age. According to Thoeny (pers. comm., 1984), a lepidopteran gall former on black locust attacks only juvenile plants and plant parts. The gall forming sawfly, *Euura lasiolepis* Smith, attacks the fastest growing shoots of arroyo willow (Craig et al., 1986) and water stress causes heavy sawfly mortality, due, in part, to its negative effects on plant vigor (Price, 1988; Price and Clancy, 1986). The bud galler, *Euura mucronata* (Hartig) Man. (Churchill), also attacks younger, more vigorously growing plants (Price et al., 1987a, b).

We studied the relationship between physical and chemical plant traits in vigorous and stressed willow plants and their effects on an associated herbivore. Our major aim was to find the plant character which best predicts sawfly attack and survivorship, using simple chemical tests of protein and phenol levels and measurement of a physical character, plant growth.

The arroyo willow, Salix lasiolepis Bentham, and a shoot galling sawfly specific to this willow, Euura lasiolepis Smith (Hymenoptera: Tenthredinidae) were examined. The willow grows as a shrub up to 4 meters tall, forming clones of a few m² to 32 m² in size. Aspects of clonal dynamics, sawfly attack and sawfly life history have been reported (Price and Craig, 1984; Price, 1988; Price and Clancy, 1986; Craig et al., 1986).

We analyzed phenolic glycosides in this study because they are the chemicals most strongly implicated in poplar and willow (Salicaceae) defense (Rowell-Rahier, 1984; Palo, 1984; Tahvanainen et al., 1985a, b; Markham, 1971; Pasteels et al., 1983; Smiley et al., 1985; Rowell-Rahier and Pasteels, 1982). Zucker (1982) suggested their possible importance in the defense of narrowleaf cottonwood (*Populus angustifolia* James) against a gall forming aphid, *Pemphigus betae* Doane and some generalists only use willow species without these defensive compounds (Rowell-Rahier, 1984). Phenol glycoside concentrations are known to vary with-

in willow (Horn, 1985) and poplar (Zucker, 1982) populations, and may be responsible for differential attack of plants (Similey et al., 1985).

The term "plant stress" is used here in the conventional sense (e.g. Boyer, 1982; Osmond et al., 1987) as a reduction in growth or reproductive potential from that achieved under nonlimiting conditions. In our study stress was estimated by growth relative to the best growth achieved in each experiment.

## Methods

The study site was located on Museum of Northern Arizona property, near Flagstaff, Arizona (elevation 2,132 m). The two experiments were conducted at Coyote Spring, a perennial spring, with two large willows that are heavily attacked by sawflies. The native willows we used for cuttings for experiments and gall samples were located at 2 sites on Schultz Creek: Museum of Northern Arizona (MNA) and at Northland Press (NP) at the southern end of museum property.

The effects of water and fertilizer treatments on willow growth and on sawfly attack and survivorship were tested experimentally. Six willow clones were used in experiments (MNA Nos. 1–4 and NP Nos. 8, 9), representing clones which consistently sustained high or low sawfly attack (High attack: MNA 2 and 3, NP 8; Low attack: MNA 1 and 4, NP 9). One-year-old shoots were taken from each clone, rooted and planted in 18.9 litre pots filled with alluvial silty sand from MNA and NP sites. Each pot contained 4 shoots from 4 different clones. For the water treatment experiment, clones MNA Nos. 1 and 2 and NP Nos. 8 and 9 were used in each pot; for the fertilizer treatment, clones MNA No.s 1–4 were used. Plants were maintained outdoors through the summer with water applications but no fertilizer and buried in soil in October for the winter.

Potted plants were placed on platforms at the Coyote Spring site on 30 May, 1983, to intermingle with the upper canopy of the heavily attacked native clones CS Nos. 1 and 2, to expose them to colonizing sawflies. Pots were rotated every 14 days to minimize position effects on attack.

The water treatment experiment used three pots per treatment, and three treatment levels, with treatments applied between 28 May and 10 October, 1983. Springwater was applied until dishes under the pots overflowed. The low water treatment (LW) pots received water every 22 days until June 19, and every 14 days thereafter, medium water treatment (MW) pots received water every 7 days and high water treatment (HW) pots received water everyday. No fertilizer was used in these experiments.

The fertilizer experiment used 3 pots per treatment and 3 treatment levels. All pots received water daily. A fertilizer application consisted of 6.2 g Miracle Gro® dissolved in 1 liter of water. The no fertilizer treatment (OF) pots received only water, the medium treatment (LF) pots received fertilizer once per week, and the high fertilizer treatment (HF) pots received the treatment 2 times per week between 28 May and 27 June, 1983.

The number of attacks or galls per shoot was counted, and shoot length was measured on the 10 most distal shoots per plant, plus any attacked shoots on 6 July, 1983. Craig et al. (1986) have shown that gallformer colonization of willow shoots occurs at nodes and increased densities of galls on longer shoot reflect increased gall densities at individual nodes on shoots. Because the number of nodes per shoot decreases with increasing shoot length,

gallformers are not responding to an increase in oviposition sites, but rather some other feature of long shoots (Craig et al. 1986). For convenience we used number of galls per shoot as a measure of gall densities. Shoot lengths were measures of plant growth in this study. Galls were collected in December, when sawfly larvae had spun cocoons, and dissected and survivors counted (survivorship = surviving to the cocooned larval stage).

The protein and phenol concentrations of leaves were also measured to determine if they were good correlates of sawfly attack and survivorship. Leaves were sampled because they are used by sawflies in initial testing for oviposition sites and petioles of young leaves are used as oviposition sites (Price and Craig, 1984). Five young, expanded leaves were taken from 2 plants per clone per treatment in the water treatment experiment (120 samples total), and from all plants in the fertilizer treatment experiment (180 samples total) on 17 August, 1983.

Protein and phenol concentrations were also measured in developed galls and ungalled stems from wild clones. Ten galls and 10 ungalled shoots at the position at which a gall would occur, were collected from 1 clone (CS 1) in August, 1982, and each was pooled; and 15 galls and 15 ungalled shoots per clone were collected from clones MNA 2 and 3 in March, 1985, and were not pooled. The parenchymatous nutritive tissue in galls and analogous cortex tissue from ungalled shoots were extracted and analysed chemically.

Total phenols were determined using the Folin-Denis procedure as described by Zucker (1982). Since this time, Julkunen-Tiitto (1985) has recommended use of the Folin-Ciocalteu phenol reagent, however we regard our results as valid because differences between the techniques are small. Phenol concentrations are expressed as total phenol concentrations per optical density (OD) at 725 nm per mg of dry mass of plant tissue (OD<sub>725</sub>).

For protein analysis, dried plant material was extracted in 1.0 ml 0.1 N NaOH in a closed tube for 3 hours, stirred vigorously for 5 minutes, set aside for an additional 3 hours, stirred for 5 minutes and left overnight. The samples were stirred for 5 minutes and then centrifuged for 3 minutes. One tenth of a ml (0.1 ml) of this supernatant was added to a test tube along with 4.9 ml of Bio-Rad® protein dye reagent prepared according to specifications. The OD (optical density) of the sample was measured at 595 nm. Bovine gamma globulin was used as a standard and its assay was linear over a 5-fold range of concentrations. Protein concentrations were expressed as percent protein in dry weight of sample.

Principal components analysis (PCA) was used to reduce dimensionality and establish overall differences due to treatments (Afifi and Clark, 1984). PCA transformed all response variables (protein and phenol concentrations, plant growth, sawfly attack) into a single variable, the first PCA axis. A split plot ANOVA was used to test for differences in the scores along the first principal component among treatments and clones (Cochran and Cox, 1957). Following this, differences in each response variable among treatments and clones were determined using split plot ANOVA. The pot represented the experimental unit or plot and each was split by the presence of 4 clones per pot; treatments and clones were the main effects. Stepwise multiple regression (Hull and Nie, 1981) was used to measure the correlation between sawfly attack and the plant variables: shoot length (growth), protein concentrations, phenol concentrations, clone and treatment. The stepwise program enters variables in descending order of importance. Sawfly survivorship data was arcsine-square root transformed and analyzed with a oneway ANOVA (sample sizes were too small to examine clonal effects on survivorship) (Zar, 1984). Data from watering and fertilization experiments were not compared because different clones were used in the two experiments.

#### Results

# Water treatment experiment

Water treatments produced the following effects on 36 experimental plants. Split-plot ANOVA on the scores along the first principal component (all response variables transformed into one variable) indicated significant differences due to treatments ( $F_{2,3} = 18.0$ , p < .005) and clones ( $F_{3,9} = 10.00$ , p < .01).

When the variables were analyzed separately, protein concentrations decreased significantly with increasing water treatment, while phenol concentrations did not vary significantly with treatment level (Table 1). The increased concentration of protein in leaves of LW plants is a commonly observed plant response to water stress (White, 1984). Shoot lengths increased significantly with increased water treatment, indicating that water increased growth rate. The attack rate or number of galls on plants increased significantly with increased water treatment (Table 1). Multiple regression indicated that, of all variables measured, sawfly attack was most strongly and positively correlated with plant growth, while no other factors contributed significantly to the regression equation (Table 2). Increased gall densities on faster growing shoots have been shown to reflect greater densities at shoot nodes and number of nodes do not increase with shoot length (Craig et al., 1986). Significant differences occurred between clones in each analysis.

Survivorship of *E. lasiolepis* larvae did not differ significantly among water levels. There was, however, a slight trend of improved survivorship with increased water (Fig. 1A), which corresponds to host plant selection patterns of adult sawflies and plant growth patterns.

The overall pattern in this experiment was one of increased sawfly attacks on rapidly growing, well watered plants, although water stressed plants had higher protein concentrations. This suggests that growth rates of willows, rather than nutrient availability, represent the best index of a suitable resource for gall forming *E. lasiolepis*.

Table 1. Split plot ANOVA results of differences in phenol and protein concentrations, plant growth and saw-fly attack among water treatments (Mean $\pm 1$  S.E.).

	Water treat	ment level:		Probability:
Response variable	LW	MW	HW	$p<(\mathrm{df})$
% total protein	11.89 (0.27)	9.70 (0.36)	8.71 (0.30)	0.025 (2, 3)
Total phenols	0.48 (0.05)	0.47 (0.04)	0.51 (0.07)	n.s. (2, 3)
Shoot length (cm)	35.30 (3.21)	47.0 (3.75)	67.30 (7.68)	0.010 (2, 6)
Sawfly attack (mean no. of galls/shoot)	2.0 (0.72)	2.0 (0.70)	6.10 (1.84)	0.050 (2, 6)

Table 2. Multiple stepwise regression equation correlating sawfly attack with protein and phenol concentrations, shoot length (growth), clone and treatment level in water experiments (df = 5, 17). Beta or slope  $(\beta)$ , standard error of beta (S.E. of  $\beta$ ), t test testing difference of slope from 0 (t), the significance of t (sign. of t) and the change in  $\mathbb{R}^2$  with addition of each variable to the regression equation are presented.

Step variable:	β	S.E. of $\beta$	t	Sign. t	Change in R <sup>2</sup>
Shoot length	0.173	0.067	2.635	0.017	0.46
Phenols	15.300	7.978	1.918	0.072	0.51
Clone	-0.661	0.989	-0.668	0.513	0.55
Protein	1.120	1.310	0.856	0.404	0.57
Treatment	1.003	2.171	0.462 -	0.650	0.57

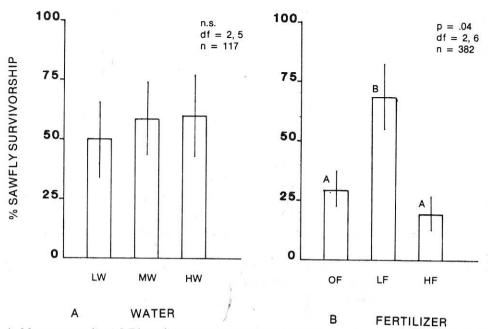


Fig. 1. Mean percent ( $\bar{x}\pm 1$  S.E.) sawfly survivorship by treatment per A. water experiment and B. fertilizer experiment.

# Fertilizer treatment experiment

Fertilizer treatments produced the following effects on 36 experimental plants. Split-plot ANOVA on the scores of the first principal component indicated significant differences due to treatments ( $F_{2,6} = 38.43, p < .005$ ) and clones ( $F_{3,18} = 5.75, p < .05$ ).

When analyzed separately, protein concentrations increased, although nonsignificantly, with increased fertilizer treatment, while phenol concentrations decreased significantly with an increase in fertilizer (Table 3). Protein and phenol concentrations were negatively correlated in this experiment ( $r^2 = .41$ , p < .05, n = 9). Unlike protein concentrations, shoot length or growth was significantly greater on LF or intermediate treatment plants, while it declined in plants receiving the most fertilizer (HF) (Table 3). Attack rates were not significantly different among treatments, although there was a slight trend of more galls occurring on plants receiving the intermediate fertilizer treatment (Table 3). Multiple regression determined that sawfly attack was most strongly correlated with plant growth, while no other variable contributed significantly to the regression equation (Table 4). Clonal differences were significant in each of these analyses.

Table 3. Split plot ANOVA results of differences in phenol and protein concentrations, plant growth and sawfly attack among fertilizer treatments (Mean $\pm 1$  S.E.)

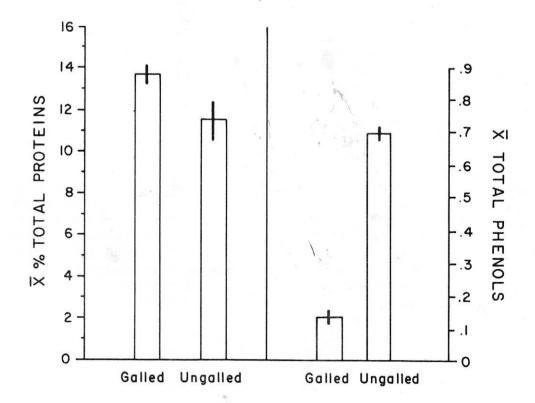
	Fer	tilizer treatment le	vel	Probability
Response variable	OF	LF	HF	p < (df)
% total proteins	9.20 (0.26)	12.34 (0.59)	13.79 (0.99)	n.s. (2, 6)
Total phenols	0.45 (0.02)	0.31 (0.02)	0.20 (0.01)	0.005 (2, 6)
Shoot length (cm)	66.80 (8.60)	158.30 (9.38)	96.30 (8.50)	0.005 (2, 6)
Sawfly attack	6.83 (2.50)	14.00 (2.50)	12.42 (3.80)	n.s. (2, 6)
(mean no. of galls/shoot)		- 2	3 2	(-, -)

Table 4. Multiple regression equation correlating sawfly attack with protein and phenol concentrations, shoot length (growth), clone and treatment level in fertilizer experiments (df = 5, 28). Beta or slope  $(\beta)$ , standard error of beta (S.E. of  $\beta$ ), t test testing difference of slope from 0 (t), significance of t (sign. of t), and the change in  $\mathbb{R}^2$  with addition of each variable to the regression equation are presented.

Step variable:	β	S.E. of $\beta$	. t	Sign. t	Change in R <sup>2</sup>
Shoot length	0.140	0.028	4.858	0.000	0.42
Clone	-2.770	1.259	-2.199	0.036	0.50
Treatment	1.986	3.289	0.604	0.551	0.50
Protein	-0.286	0.613	-0.466	0.645	0.51
Phenols	6.292	21.577	0.721	0.971	0.51

Percent survivorship of sawfly larvae was significantly greater on LF plants (Fig. 1B), which also had the longest shoot lengths. Shoot growth appears to affect the survival of sawfly eggs (Price and Clancy, 1986) and newly hatched larvae (Preszler and Price,1988). The percentage of survivorship between treatments did not differ after young larvae were established in the gall.

The results of this experiment also indicate that sawfly survivorship and to a certain extent, attack, are increased on rapidly growing plants, regardless of their biochemical status, as we have measured it.



# TISSUE SAMPLED

Fig. 2. Mean ( $\pm 1$  S.E.) protein and phenol concentrations in *E. lasiolepis* gall tissue and in ungalled tissue. Percent total soluble protein/mg dry weight plant tissue: p < 0.04, df = 1, 47; total phenol concentration (OD<sub>725</sub>): p < 0.001, df = 1, 47.

## Euura gall chemistry

Galled tissue had greater concentrations of protein per mg of dried tissue than did ungalled cortex tissue collected in August, 1982 (galled = 11.0, ungalled = 8.05). The same pattern was found for galled and ungalled tissue collected in March, 1985 and was significant (galled  $\bar{x} = 13.7\%$ , ungalled  $\bar{x} = 11.6\%$ , Fig. 2).

Phenol concentrations were lower in galled than in ungalled samples collected in August, 1982 (galled = .220, ungalled = .282). The pattern was the same and significant for galled and ungalled samples collected in March, 1985 (galled  $\bar{x} = .140$ , ungalled  $\bar{x} = .698$ , Fig. 2). We propose that the resource is being improved by this herbivore and the gall forming process. This may explain the lack of correlation between gall former host selection and survivorship and concentrations of these chemicals in the experiments.

# Discussion

Experimental treatments resulted in a complex mosaic of physical and biochemical conditions in the willows, with the relationship between nutrition, defense and vigor dependent upon the nature and intensity of each treatment. Our experiments decoupled the relationships of plant growth, and phenol and protein concentrations to one another. In the midst of this complexity, however, *Euura lasiolepis* responded strongly and positively to the physical condition of vigor.

Plant chemistry, as measured by total phenol and protein concentrations, was not a useful index or correlate of colonization or survivorship patterns of *E. lasiolepis* in this study. Although plants stressed from lack of water or excessive fertilizer exhibited the highest concentrations of total soluble proteins, colonization (water experiment) or survivorship (fertilizer experiment) of sawflies was greatest on plants without the highest protein levels. These findings run counter to the prediction of White (1984) that stressed plants provide better resources for colonizing herbivores, particularly early instars, because in both of our experiments attack rates and/or survivorship were greatest on the least stressed plants, with stress negatively affecting early instar sawfly survivorship.

Phenolic levels were also poor predictors of *E. lasiolepis* attack or survival. If phenols were functioning defensively, a negative correlation between phenols and attack or survivorship should have been observed. Interestingly, plant protein and phenol levels were inversely correlated. High protein concentrations, combined with low phenol concentrations can actually reduce the toxicity of phenolic compounds (Kirkham, 1954; Kiraly, 1976), seemingly rendering such plants ideal resources for herbivores. Despite our findings, we recognize that individual phenolic glycosides may play a role in host plant selection, as determined for other herbivores (Pasteels et al., 1983; Tahvanainen et al., 1985b; Smiley et al., 1985) and this possibility will be tested in the future.

Euura lasiolepis' preference for fast growing plant tissue, with its counterparts in other gall forming and free feeding species of herbivores, emphasizes the need to understand the nature of growth itself and precisely what it has to offer herbivores. Ellis et al. (1965) found that the desert locust could not attain sexual maturity when fed senescent plant tissue. Growth hormones, abundant in growing tissue, may function in gall development (Arrillaga, 1949; Balch et al., 1964; Bouillenne and Gaspar, 1970). Byers et al. (1976) found that levels of the growth hormones, gibberillins and auxins, were respectively 21 and 17 times more abundant in pinyon galls than ungalled tissue formed by the cecidomyiid, Janetiella sp., and low or absent in the midge larvae themselves.

We hypothesize that *E. lasiolepis* may be modifying willow tissue locally, because protein concentrations were greater and phenolic concentrations lower in galled tissue, which may explain the lack of correlation between these compounds and sawfly attack and survivorship. Washburn (1984) has presented evidence of gall former resource modification for natural enemy defense on oaks, and evidence for resource modification for nutrition exists in other gallforming systems: Braun (1969), Waring (unpublished data) and Abrahamson and McCrea (1986) found nitrogen and/or protein levels to be greater in galled than ungalled tissue in three separate systems; Bronner (1976), Meyer (1954) and Rey et al. (1980) each found higher lipid concentrations in galled versus ungalled tissue in three separate studies and Shannon and Brewer (1980) found starch concentrations to be greater in galled tissue in three separate systems (see also Abrahamson and Weis, 1987).

This study emphasizes the need to modify the developing theory on plant quality and herbivores. Many herbivores respond positively to vigorously growing members of a plant population, as in *Euura lasiolepis* and *E. mucronata* (Price et al., 1987a, b). So a dichotomy must be recognized: the vigorous plant herbivores and the stressed plant herbivores, perhaps with a continuum between them. Also, stress is not a simple phenomenon, but complicated by distinct factors such as water or nutrients, within each of which there is a continuum of responses by the plant and the herbivores. In our experiments water stress increased protein content, without affecting phenol content, while relieving nutrient stress with the low fertilizer treatment caused increased protein and reduced phenol concentrations. Thus, in the future, stress must be defined in specific terms relative to known response curves of herbivores to changing concentrations of nutrients and defensive chemicals. Although the theory cannot be as general as we would like, neither can it be a mechanistic theory without recognizing the details of plant responses to changing physical factors, and herbivore responses to plant variation under stress.

# Acknowledgments

We thank M. Rowell-Rahier, L. Stevens, J. Tahvanainen and W. Zucker for their critical comments on a draft of this paper. Thanks to W. Zucker and P. Evans for assistance with biochemical methodology, and to B. Boecklen and G. Bell for assistance with statistical design. E. L. Smith of the California Academy of Sciences identified *Euura lasiolepis* and A. Phillips III of the Museum of Northern Arizona, identified *Salix lasiolepis*. Financial support for this study was provided by N. S. F. grants DEB-8021754 and BSR-8314594.

#### References

ABRAHAMSON, W. G. & McCrea, J. D. (1986) Nutrient and biomass allocation in *Solidago altissima*: effects of two stem gallmakers, fertilization, and ramet isolation, Oecologia **68**: 174–180.

——— & Weis, A. E. (1987) Nutritional ecology of arthropod gallmakers. "Nutritional ecology of insects, mites, spiders and related invertebrates" (ed. Slansky, F. & Rodriguez, J. G.), 235–258. Wiley Interscience, New York.

AFIFI, A. A. & CLARK, V. (1984) Computer-aided multivariate analysis. Wadsworth, Inc. London. ARRILLAGA, J. G. (1949) Formation of galls in stems and leaves in sugarcane in response to injections of growth-regulating substances. Phytopathology 39: 489–493.

Askew, R. R. (1962) The distribution of galls of *Neuroterus* (Hym.: Cynipidae) on oak. J. Anim. Ecol. 31: 439-455.

BAKKE, A. (1969) The effect of forest fertilization on the larval weight and larval density of Laspeyresia

- strobilella (L.) (Lepidoptera: Tortricidae) in cones of Norway spruce. Zeitschrift für Angewandte Entomologie 63: 451–453.
- BALCH, R. E., CLARK, J. & BONGA J. M. (1964) Hormonal action in production of tumors and compression wood by an aphid. Nature 202: 721-722.
- Berryman, A. A. (1982) Population dynamics of bark beetles. "Bark Beetles in North American conifers" (ed. Mitton, J. B. & Sturgeon, K. B.), 265–314. University of Texas Press, Austin.
- BOUILLENNE, C. & GASPAR, T. (1970) Auxin catabolism and inhibitors of normal and crown gall tissues of *Impatiens balsamina* Can. J. Bot. 48: 1159-1163.
- BOYER, J. S. (1982) Plant productivity and environment. Science 218: 443-448.
- Braun, A. C. (1969) Abnormal growth in plants. "Plant physiology: a treatise, vol. VB. Analysis of growth: the responses of cells and tissues in culture" (ed. Steward F.C.), Academic Press, New York.
- Bronner, R. (1976) Contribution a l'étude histochimique des tissus nourriciers des zoocecidies. Strasbourg: These.
- Byers, J. A., Brewer, J. W. & Denna, D. W. (1976) Plant growth hormones in pinyon insect galls. Marcellia 39: 125-134.
- CATES, R. G., REDAK, R. A. & HENDERSON, C. B. (1983) Patterns in defensive natural product chemistry: Douglas fir and western spruce budworm interactions. "Plant resistance to insects" (ed. Hedin P. A.), 3–9. Amer Chem Soc, Washington, D.C.
- CHAPMAN, R. F. & BERNAYS, E. A. (1978) Insect and host plant. Entomol. Exp. Appl. 24: 1–566.
- COCHRAN, W. G. & Cox, G. M. (1957) Experimental designs. John Wiley & Sons, New York.
- CRAIG, T. P., PRICE, P. W. & ITAMI, J. K. (1986) Resource regulation by a stem-galling sawfly on the arroyo willow. Ecology 67: 419-425.
- EHRLICH, P. R. & RAVEN, P. H. (1964) Butterflies and plants: A study in coevolution. Evolution 18: 586-608.
- ELLIS, P. E., CARLISLE, D. B. & OSBORNE, D. J. (1965) Desert locusts: sexual maturation delayed by feeding on senescent vegetation. Science 149: 546-547.
- FRAENKEL, G. (1959) The raison d'etre of secondary plant substances. Science 129: 1466-1470.
- ——. (1969) Evaluation of our thoughts on secondary plant substances. Entomol. Exp. Appl. 12: 473–486.
- Frankie, G. W. & Morgan, D. L. (1984) Role of the host plants and parasites in regulating insect herbivore abundance, with an emphasis on gall-inducing insects. "A new ecology: Novel approaches to interactive systems" (ed. Price, P. W., Slobodchikoff, C. N. & Gaud, W. S.), 101–104. Wiley, New York,
- HARBORNE, J. B. (ed.) (1972) Phytochemical ecology. Academic Press, New York.
- ———. (1977) Introduction to ecological biochemistry. Academic Press, New York.
- ——. (ed.) (1978) Biochemical aspects of plant and animal coevolution. Academic Press, New York.
- HAWKSWORTH, F. G. (1960) Growth rate of dwarf mistletoe infections in relation to the crown class of the host. Rocky Mtn For Range Exp Stat Res Notes 41: 1-4.
- HEDIN, P. A. (ed.) (1983) Plant resistance to insects. Amer Chem Soc, Washington, D.C.
- HORN, J. M. (1985) Distribution of phenolglycosides in willow (Salix spp.) leaves. White Mtn Res Stat Symp Proc 1985: 1–18.
- HUFFAKER, C. B., VAN DE VRIE, M. & MCMURTY, J. A. (1969) The ecology of tetranychid mites and their natural control. Ann. Rev. Ent. 14: 125–174.
- HULL, C. H. & NIE, N. H. (1981) SPSS-Update. McGraw-Hill, Inc. NY.
- HUSSEY, N. W. & HUFFAKER, C. B. (1976) Spider mites. "Studies in biological control" (ed. Delucchi, L. V.), 179–228. Cambridge Univ. Press, London.
- JULKUNEN-TIITO, R. (1985) Phenolic constituents in the leaves of northern willows: Methods for the analysis of certain phenolics. J. Agr. Food Chem. 33: 213–217.
- KIRALY, Z. (1976) Plant disease resistance as influenced by biochemical effects of nutrients in fertilizers. "Fertilizer use and plant health" (ed. Dimitri & Bogenschutz), Int. Potash Inst. Worblaufen-Bern, Switzerland. pp. 33-45.
- KIRKHAM, D. S. (1954) Significance of the ratio of the water soluble aromatic and nitrogen constituents of apple and pear in the host-parasite relationship of *Venturia* sp. Nature 137: 690-691.
- MARKHAM, K. R. (1971) A chemotaxonomic approach to the selection of oppossum resistant willows and poplars for use in soil conservation. New Zealand J. Sci. 14: 179–186.

- MEYER, J. (1954) Sur l'accumulation et l'origine de lipides dans certains tissus nourriciers de galles. C. R. Acad. Sci. Paris 238: 1066–1069.
- ONUFF, C. P., TEAL, J. P. & VALIE, I. (1977) Interactions of nutrients, plant growth and herbivory in a mangrove ecosystem. Ecology 58: 514–526.
- OSMOND, C. B., AUSTIN, M. P., BERRY, J. A., BISSONGS, W. D., DACEY, J., NOBEL, P. S., SMITH, S. D. & WINNER, W. E. (1987) Stress physiology and the distribution of plants. BioScience 37: 38–48.
- Palo, R. T. (1984) Distribution of birch (*Betula* spp.) and willow (*Salix* spp.), and poplar (*Populus* spp.) secondary metabolites and their potential role as chemical defenses against herbivores. J. Chem. Ecol. 10: 499–520.
- Pastells, J. M., Rowell-Rahier, M., Braekman, J. C. & Dupont, A. (1983) Salicin from host plant as precursor of salicylaldehyde in defensive secretion of chrysomeline larvae. Phys. Ent. 8: 307–314.
- Preszler, R. W. & Price, P. W. (1988) The effects of variable host quality on sawfly populations; A new approach to life table analysis. Ecology (In press).
- PRICE, P. W. (1988) Biology of insect and mite induced galls. "Evolution and ecology of gall-inducing sawflies" (ed. Shorthouse, J. D. & Rohfritsch, O.) Preager, New York (In press).
- ——— & Craig, T. P. (1984) Life history, phenology and survivorship of a stem-galling sawfly, Euura lasiolepis (Hymenoptera: Tenthredinidae), on the arroyo willow, Salix lasiolepis, in Northern Arizona. Ann. Ent. Soc. Amer. 77: 712–719.
- ——— & CLANCY, K. M. (1986) Multiple effects of precipitation on *Salix lasiolepis* and populations of the stem-galling sawfly, *Euura lasiolepis*. Ecol. Res. 1: 1–14.
- ———, ROININEN, H. & TAHVANAINEN, J. (1987a) Plant age and attack by the bud galler, *Euura mucronata*. Oecologia 73: 334–337.
- REY, L., DUBACQ, J. P. & TREMOLIERES, A. (1980) Lipids of oak galls. Phytochem. 19: 2569-2570.
- RHOADES, D. F. (1979) Evolution of plant chemical defense against herbivores. "Herbivores: Their interaction with secondary plant metabolites" (ed. Rosenthal, G. A. & Janzen, D. H.), 3–54. Academic Press, New York.
- ———. (1983) Herbivore population dynamics and plant chemistry. "Variable plants and herbivores in natural and managed systems" (ed. Denno, R. F. & McClure, M. S.), 155–220. Academic Press, New York.
- ROSENTHAL, G. A. & JANZEN, D. H. (eds.) (1979) Herbivores: Their interaction with secondary plant metabolites. Academic Press, New York.
- ROWELL-RAHIER, M. (1984) The presence or absence of phenol glycosides in *Salix* (Salicaceae) leaves and the level of dietary specialization of some of their herbivorous insects, Oecologia **62**: 26–30.
- & Pastells, J. M. (1982) The significance of salicin for a *Salix*-feeder, *Phatora* (*Phyllodecta*) vitellinae. "Insect-plant relationships" (ed. Visser, J. H. & Minks, A. K.), 73–79. Center for Agric. Publ. Doc., Wageningen, The Netherlands.
- Shannon, R. E. & Brewer, J. W. (1980) Starch and sugar levels in three coniferous insect galls. Zeitschrift für angewandte Entomologie 5: 526–533.
- SHAW, G. G. & LITTLE, C. H. A. (1972) Effect of high urea fertilization of balsam trees on spruce budworm development. "Insect and mite nutrition" (ed. Rodriguez, J. G.), 589–597. North Holland, Amsterdam, The Netherlands.
- SMILEY, J. T., HORN, J. M. & RANK, N. E. (1985) Ecological effects of salicin at three trophic levels: new problems from old adaptations. Science 229: 649-651.
- Tahvanainen, J., Helle, E., Julkunen-Tiitto, R. & Lavola, A. (1985a) Phenolic compounds of willow bark as deterrents against feeding by mountain hare. Oecologia 63: 319–323.
- ———, Julkunen-Tiitto, R. & Kettunen, J. (1985b) Phenolic glycosides govern the food selection pattern of willow feeding leaf beetles. Oecologia 67: 52–56.
- VAN EMDEN, H. F., EASTOP, V. F., HUGHES, R. D. & WAY, M. J. (1969) The ecology of Myzus persicae. Ann. Rev. Ent. 14: 197–270.
- VISSER, J. H. & MINKS, A. K. (eds.) (1982) Insect-plant relationships. Center for Agric. Publ. Document, Wageningen, The Netherlands.
- Wallace, J. W. & Mansell, R. L. (eds.) (1976) Biochemical interaction between plants and insects. Plenum Press, New York.
- WASHBURN, J. O. (1984) Mutualism between a cynipid gall wasp and ants. Ecology 65: 654-656.
- ——— & CORNELL, H. V. (1981) Parasitoids, patches and phenology: Their possible role in the local extinction of a cynipid gall wasp population. Ecology **62**: 1597–1607.

Weis, A. E., Walton, R. & Crego, C. L. 1988. Reactive plant tissue sites and the population biology of gall makers. Ann. Rev. Entomol. 33: 467–486.

WHITE, T. C. R. (1974) A hypothesis to explain outbreaks of looper caterpillars, with special reference to populations of *Selicosema suavis* in a plantation of *Pinus radiata* in New Zealand. Oecologia 16: 279–301.

—. (1976) Weather, food and plagues of locusts. Oecologia 22: 119–134.

... (1978) The importance of relative shortage of food in animal ecology. Oecologia 33: 71–86.
... (1984) The abundance of invertebrate herbivores in relation to the availability of nitrogen in stressed food plants. Oecologia 63: 90–105.

ZAR, J. H. (1984) Biostatistical analysis. Prentice-Hall, N.J.

Zucker, W. V. (1982) How aphids choose leaves: The role of phenolics in host selection by a galling aphid. Ecology 63: 972–981.